# ESTIMATING SPECIES TREES USING MULTIPLE-ALLELE DNA SEQUENCE DATA 

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#### Abstract

Several techniques, such as concatenation and consensus methods, are available for combining data from multiple loci to produce a single statement of phylogenetic relationships. However, when multiple alleles are sampled from individual species, it becomes more challenging to estimate relationships at the level of species, either because concatenation becomes inappropriate due to conflicts among individual gene trees, or because the species from which multiple alleles have been sampled may not form monophyletic groups in the estimated tree. We propose a Bayesian hierarchical model to reconstruct species trees from multipleallele, multilocus sequence data, building on a recently proposed method for estimating species trees from single allele multilocus data. A two-step Markov Chain Monte Carlo (MCMC) algorithm is adopted to estimate the posterior distribution of the species tree. The model is applied to estimate the posterior distribution of species trees for two multiple-allele datasets-yeast (Saccharomyces) and birds (Manacus—manakins). The estimates of the species trees using our method are consistent with those inferred from other methods and genetic markers, but in contrast to other species tree methods, it provides credible regions for the species tree. The Bayesian approach described here provides a powerful framework for statistical testing and integration of population genetics and phylogenetics.


KEY WORDS: Bayesian hierarchical model, coalescent theory, gene tree, species tree.

The advance of molecular biological technologies has enabled the rapid collection of multiple alleles from a given species in the context of building phylogenetic trees of multiple species. Multiple-allele data contain more information about the evolutionary history of species than single allele data, and recent years have seen a growth of statistical approaches that effectively combine the multiple-allele information from multiple loci. These

[^0]approaches address several important phylogeographic models, such as gene flow, population growth, and population divergence (Nielsen and Slatkin 2000; Beaumont and Rannala 2004; Hey and Nielsen 2004; Degnan and Salter 2005; Beerli 2006; Degnan and Rosenberg 2006). However, available techniques for estimating phylogenetic trees from multiple-allele data remain limited, despite the fundamental importance of trees for historical inference. Commonly used techniques, such as the concatenation method (Nylander et al. 2004), the consensus tree method (Bull et al. 1993; de Queiroz 1993; Huelsenbeck et al. 1994; de Queiroz et al.

1995; Wiens 1998), and gene tree parsimony (Page and Charleston 1997; Slowinski et al. 1997; Page 1998) are not well suited for pooling the information from multiple alleles and multiple genes to build a species tree in which only the actual species, instead of all individual alleles, are of interest. These techniques may not be able to provide useful information on the ancestral history of species if the individual alleles of a species appear to be polyphyletic, which unfortunately occurs in many multipleallele datasets, even for mitochondrial DNA (Funk and Omland 2003).

The deep coalescence approach (Maddison and Knowles 2006) has been shown to be useful in estimating species trees from multiple-allele data. It does so by finding the species tree that minimizes the topological discrepancy-the number of deep coalescences-between the collected gene trees and the proposed tree, summed over all genes. Although useful, this approach ignores information about branch lengths in gene trees, which may compromise and limit the utility of phylogenetic inference. A Bayesian hierarchical model has been proposed to estimate species trees for single allele data (Edwards et al. 2007; Liu and Pearl 2007). In this article, we extend that Bayesian hierarchical model to reconstruct species trees from multiple-allele data. The multiple-allele Bayesian hierarchical model is able to extract information from all individual alleles of a species to make inferences concerning the ancestral history of the species and therefore estimate a species tree, despite the fact that the taxonomic units in the gene trees are alleles. Using simulations, we show that the Bayesian estimate appears to be statistically consistent in the sense that it moves closer to the true species tree in probability as the number of genes and the sequence length go to infinity, even in situations in which there is substantial topological and branch length heterogeneity among genes and between the gene tree and species tree.

## Details of the Model

We use the following abbreviations: D: Sequence data; $\mathbf{G}$ : a vector of gene trees across genes; $\Lambda$ : Parameters in the likelihood function except the gene tree vector $\mathbf{G} ; \mathrm{S}$ : Species tree; $\theta$ : Transformed effective population sizes, $\theta=4 N_{e} \mu$.

The Bayesian hierarchical model consists of five components: (1) the likelihood function, $f(D \mid \mathbf{G}, \Lambda)$; (2) the prior distribution of $\Lambda, f(\Lambda)$; (3) the probability distribution of gene tree vector $\mathbf{G}$ given the species tree S (topology and branch lengths) and $\theta, f(\mathbf{G} \mid S, \theta)$; (4) the prior distribution of the species tree, $f(S)$; and (5) the prior distribution of $\theta, f(\theta)$.

The likelihood $f(D \mid \mathbf{G}, \Lambda)$ is derived from substitution models such as the HKY (Hasegawa et al. 1985) or GTR (Lanave et al. 1984) model for nucleotides or the WAG model for proteins (Whelan and Goldman 2001). The prior distribution of $\Lambda$ depends


Figure 1. A multiple-allele gene tree and a species tree. There are two alleles for species $A\left(A_{1}\right.$ and $\left.A_{2}\right)$, three alleles for species $B$ ( $B_{1}, B_{2}, B_{3}$ ), and one allele for species $C\left(C_{1}\right)$. The shaded regions in the gene tree represent the corresponding populations in the species tree. Populations $A$ and $B$ are contemporary populations for species $A$ and $B$. Population $A B$ is the ancestral population of species $A$ and $B$. Population $A B C$ is the ancestral population of species $A, B$, and $C$.
on the nature of the data at hand. Different users may choose different priors for $\Lambda$. For example, a variety of options for the prior of $\Lambda$ are available in MrBayes (Huelsenbeck and Ronquist 2001; Ronquist and Huelsenbeck 2003), which we use to sample gene trees from the posterior distribution.

The probability distribution of gene trees given the species tree has been derived under the Kingman coalescent process (Kingman 1982, 2000) for the case in which multiple alleles are sampled from individual species related in a phylogeny (Rannala and Yang 2003). For example, consider the case when there are three species-A, B, and C (Fig. 1). In this example, two gene copies are sampled from species A, three copies from species B, and one copy from species C . The shaded areas in A and B represent current populations of species $A$ and $B$. Population $A B$ is the common ancestral population of species A and B, whereas ABC is the common ancestral population of all three species. The population size of an extant population is considered only if multiple alleles are sampled from that species. In this example, species A and B have estimable population sizes.

The probability distribution of a gene tree topology and the ( $m-n$ ) coalescent times $t_{n+1}, \ldots, t_{m}$ for a single population reduced from $m$ to $n$ sampled alleles along a branch of length $\tau$ in a species tree is (Rannala and Yang 2003)

$$
\begin{equation*}
\exp \left(-\frac{n(n-1)}{\theta}\left(\tau-\sum_{j=n+1}^{m} t_{j}\right)\right) \prod_{j=n+1}^{m}\left[-\frac{2}{\theta} \exp \left(-\frac{j(j-1)}{\theta} t_{j}\right)\right] . \tag{1}
\end{equation*}
$$

For a vector of gene trees, $\mathbf{G}$, that are independent given the species tree, we multiply (1) across gene trees to find the likelihood for a single population, i.e.,

$$
\begin{align*}
\prod_{k=1}^{W}\{\exp ( & \left.-\frac{n_{k}\left(n_{k}-1\right)}{\theta}\left(\tau-\sum_{j=n_{k}+1}^{m_{k}} t_{k j}\right)\right) \\
& \left.\times \prod_{j=n_{k}+1}^{m_{k}}\left[\frac{2}{\theta} \exp \left(-\frac{j(j-1)}{\theta} t_{k j}\right)\right]\right\} \tag{2}
\end{align*}
$$

in which $W$ is the number of genes. We can simplify (2) as

$$
\begin{equation*}
\left(\frac{2}{\theta}\right)^{a} e^{\frac{-b}{\theta}} \tag{3}
\end{equation*}
$$

where $b=\sum_{k=1}^{W}\left\{n_{k}\left(n_{k}-1\right)\left(\tau-\sum_{j=n_{k}+1}^{m_{k}} t_{k j}\right)+\sum_{j=n_{k}+1}^{m_{k}}\right.$ $\left.j(j-1) t_{k j}\right\}$ and $a=\sum_{k=1}^{W}\left(m_{k}-n_{k}\right)$. We use a conjugate prior for $\theta$ to reduce the computational demand for species tree estimation (Hey and Nielsen 2007). With a conjugate prior, the parameter $\theta$ can be more easily integrated out of the likelihood function, enabling us to form a new likelihood function without $\theta$. Here we choose the inverse gamma distribution as the conjugate prior of $\theta$, i.e.,

$$
f(\theta)=\frac{\beta^{\alpha}}{\Gamma(\alpha)} \theta^{-\alpha-1} e^{-\beta / \theta}
$$

with mean $\frac{\beta}{\alpha-1}$ and variance $\frac{\beta^{2}}{(\alpha-1)^{2}(\alpha-2)}$ for $\alpha>2$. The likelihood of a gene tree in a single population is then obtained by integrating out $\theta$ in (3) with respect to the conjugate prior of $\theta$, i.e.,

$$
\int_{\theta}\left(\frac{2}{\theta}\right)^{a} e^{-\frac{b}{\theta}} \frac{\beta^{\alpha}}{\Gamma(\alpha)} \theta^{-\alpha-1} e^{\frac{-\beta}{\theta}} d \theta=\frac{2^{a} \beta^{\alpha}}{\Gamma(\alpha)} \frac{\Gamma(\alpha+a)}{(\beta+b)^{(\alpha+a)}}
$$

The probability density $f(\mathbf{G} \mid S)$ of the gene tree vector $\mathbf{G}$ given the species tree is the product of such likelihoods across all extant and ancestral populations under the assumption that the coalescent processes within different populations are independent. Apparently, $f(\mathbf{G} \mid S)$ does not involve $\theta$. When using an MCMC (Hastings 1970) algorithm to estimate the posterior distribution of the species tree, we would then not have to update $\theta$ and would therefore reduce the computational burden for our species tree estimation routine, although $\theta$ can still be estimated with this approach. The parameters $\alpha$ and $\beta$ in the inverse gamma distribution reflect the magnitude of influence of the prior distribution of population sizes on the function $f(\mathbf{G} \mid S)$. Large values of $\alpha$ and $\beta$ may result in strong influence of the inverse gamma prior on the function $f(\mathbf{G} \mid S)$. We suggest using small values (e.g., $\alpha=3$ and $\beta=0.03$ ) for $\alpha$ and $\beta$ to reduce the effect of the prior on the function $f(\mathbf{G} \mid S)$ unless there exists some information about the possible values of $\alpha$ and $\beta$. Another choice for the prior on $\theta$ is the gamma distribution. In our current implementation, the user is allowed to use the gamma distribution, but this approach is somewhat slower than when using the inverse-gamma. Both priors, however, are able to yield estimates of the posterior distribution of the parameter $\theta$ for each node in the species tree.

Finally, we assume that the prior distribution of the topology and branch lengths of the species tree follows a birth-and-death process (Nee et al. 1994; Alfaro and Holder 2006).

## Molecular Clock

Coalescent theory assumes that a molecular clock holds for gene trees. However, when estimating gene trees from DNA sequences, the assumption of a molecular clock may introduce estimation bias. To relax this assumption, an unrooted gene tree unconstrained by a molecular clock is proposed to calculate the probability of DNA sequences given the gene tree as in a typical Bayesian analysis. The proposed gene tree is then rooted by an outgroup. For computational ease, the model only accommodates a single outgroup sequence. The rooted gene tree is then converted to an ultrametric tree to calculate the probability of the gene tree given the species tree using an ad hoc method described by Edwards et al. (2007) in which all tips of the gene tree are made contemporaneous and then the total tree length is normalized to the original, nonclock tree length.

## Algorithm

The MCMC algorithm is implemented to estimate the posterior distribution of the species tree. It is unnecessary to update $\theta$ in the MCMC algorithm because $\theta$ has already been integrated out of the model and the probability distribution of gene trees only depends on the species tree (topology and branch lengths). As a consequence, updating the species tree is based on the prior distribution $f(S)$ and the probability density function $f(\mathbf{G} \mid S)$. The posterior distribution of the species tree is

$$
f(S \mid D)=\int_{\mathbf{G}} f(\mathbf{G} \mid D) f(S \mid \mathbf{G}) d \mathbf{G}
$$

i.e., the posterior of the species tree given gene trees $f(S \mid \mathbf{G})$ weighted by $f(\mathbf{G} \mid D)$. This motivates a two-step MCMC algorithm in which the posterior distribution of gene trees is estimated in the first MCMC and then used to estimate the posterior distribution of the species tree in the second MCMC (Liu and Pearl 2007).

However, in the first MCMC algorithm using DNA sequences to estimate the posterior of gene trees, the prior of gene trees, $f(\mathbf{G})$, is unknown. Theoretically, $f(\mathbf{G})$ is equal to the integration of $f(\mathbf{G} \mid S)$ with respect to the species tree (topology and branch lengths),

$$
f(\mathbf{G})=\int_{S} f(\mathbf{G} \mid S) f(S) d S
$$

We apply the harmonic mean technique (Newton et al. 1994) to approximate the joint probability distribution, $f(\mathbf{G})$, of the gene
tree vector by $\tilde{f}(\mathbf{G})=\left(\sum_{j=1}^{k} \frac{1}{f\left(\mathbf{G} \mid S_{j}\right)}\right)^{-1}$, where $\left\{S_{j}, j=1, \ldots, k\right\}$ are the species trees generated from an MCMC sampler for which we use the Maximum Tree (MT) (Liu 2006; Edwards et al. 2007; Liu and Pearl 2007; Mossel and Roch 2007) as the start tree. The MT is the tree in the species tree space with the longest possible branches that are temporally compatible with all gene trees in the vector. It has been shown that the MT is itself a consistent estimator of the species tree if the gene trees are given without error (Liu 2006; Liu and Pearl 2007; Mossel and Roch 2007). Using the MT as the start tree greatly increases the convergence rate of the MCMC sampler and reduces the computational time, in the same way that starting a typical Bayesian analysis with a neighborjoining tree, or any other good approximation of the tree, would accelerate convergence. Although we use the MT as the starting tree, the algorithm still samples an arbitrary number of species trees differing in topology and branch lengths. The accuracy of the approximation depends in part on the number of species trees sampled in this first step. However, sampling a large number of species trees will dramatically increase the computational burden of the algorithm. As a trade-off, we instead sample a relatively small number of species trees to calculate $\tilde{f}(\mathbf{G})$.

We have incorporated the approximate joint probability distribution of the gene tree vector $\tilde{f}(\mathbf{G})$ into the popular Bayesian phylogenetic program MrBayes (Huelsenbeck and Ronquist 2001). This modified version of MrBayes provides an option for users to estimate gene trees jointly as described above. In the second MCMC algorithm, the posterior distribution of the species tree is estimated for each gene tree vector generated from the first MCMC algorithm. The sample of species trees across all gene tree vectors is used as the provisional estimate of the posterior distribution of the species tree. Additionally, $\tilde{f}(\mathbf{G})$ is recalculated in the second MCMC algorithm by sampling a large number of species trees. Finally, we use importance sampling to correct the error made in calculating $\tilde{f}(\mathbf{G})$ by using an approximate prior on gene trees in the first MCMC algorithm.

The number of species trees that are sampled to calculate $\tilde{f}(\mathbf{G})$ in the first MCMC is determined by two factorscomputational burden and accuracy of the approximation. We suggest running a small pilot trial in which only a few gene tree vectors (e.g., 100 for an eight-species tree) are generated in the first MCMC and find the minimum number of species trees required to be sampled to deliver a relatively accurate approximation of $f(\mathbf{G})$, as indicated by the difference between the approximate $\tilde{f}(\mathbf{G})$ in the first MCMC algorithm and the recalculation of $\tilde{f}(\mathbf{G})$ in the second MCMC algorithm. Further work is needed to develop an algorithm to automatically find an optimal number of species trees to approximate the joint probability distribution of gene trees.

The convergence of the first MCMC algorithm is assessed by setting memcdiagn $=$ yes in MrBayes if two or more com-
pletely independent analyses have been conducted for the dataset (Huelsenbeck and Ronquist 2001; Ronquist and Huelsenbeck 2003). However, running two analyses will dramatically increase the computation time. More commonly, the convergence of the MCMC algorithm is evaluated for a single run by examining the log-likelihood values, which is also used to monitor the convergence of the second MCMC algorithm.

Any number of summaries of the posterior distribution of species trees can be used (Beaumont and Rannala 2004; Cranston and Rannala 2007). As an estimate of the species tree, we use the majority rule consensus tree of the sample of species trees generated from the algorithm. The multiple-allele algorithm has been incorporated in the program BEST (Bayesian Estimation of Species Trees, version 1.6) and is available for download (www.stat.osu.edu/ $\sim \mathrm{dkp} / \mathrm{BEST}$ ).

## Simulation

To test the multiple-allele BEST method, three species trees were generated randomly using the Yule model in Mesquite version 1.12 (Maddison and Maddison 2006) with the fixed tree height 1. The species trees had $10,20,30$ species with three alleles per species. The value of $\theta$ was generated from a uniform distribution $(0,0.1)$ for each population in the three species trees and used in MCMCcoal (Rannala and Yang 2003) to simulate 50 gene tree vectors of $5,10,20,30$, and 40 gene trees. The length of the gene tree vector represented the number of loci. The gene tree vectors were then used to estimate the posterior distribution of the species tree using BEST. For each gene tree vector, a consensus tree was constructed from the estimated posterior distribution of the species tree and used as the estimate of the species tree. No DNA sequences were used in this simulation-we used the generated gene tree vectors as data. Thus, although there was substantial gene tree heterogeneity, there was no gene tree error in these simulations. The proportion of trials yielding the true species tree is reported in Figure 2. The discrepancy between the true species tree and the consensus tree in both topology and branch lengths was measured by the branch score distance (Kuhner and Felsenstein 1994; Felsenstein 2004).

The results show that the species tree estimate given by BEST converges to the true species tree with probability 1 as the number of loci increases (Fig. 2). The convergence rate depends primarily on the number of species. The convergence rate for 30 species is much slower than that for 10 species. With five loci, the proportion of trials yielding the true species tree is 0.38 for the tree with 30 species, whereas it is 0.96 for the tree with 10 species. Similarly, the branch score distance appears to converge to 0 as the number of loci increases (Fig. 3). The convergence rate for branch score distance is negatively related to the number of loci. The results suggest that the multiple-allele BEST method (topology and


Figure 2. The proportion of trials yielding the true species tree versus the number of genes. A set of $\mathbf{5 0}$ gene tree vectors of 5 , 10, 20, 30, 40 genes were generated from three species trees simulated from Mesquite (Maddison and Maddison 2006). Each gene tree vector was used as data to estimate the species tree. The proportion of trials yielding the true species tree appears to converge to 1 as the number of loci increases, but the convergence rate is negatively correlated with the number of species.
branch length) is statistically consistent when the gene trees are given without errors.

In the second simulation, DNA sequences were generated from three prespecified species trees and used as the data to estimate the species phylogeny. The simulation was conducted for four species: A, B, C, and D. The species A, B, and C had two alleles respectively, whereas species $D$ had one sequence. The three species trees were: Tree 1: (((A:0.01, B:0.01):0.001, C:0.011):0.1, D:0.111); Tree 2: (((A:0.01, B:0.01):0.005, C: $0.015): 0.1, \mathrm{D}: 0.115)$; Tree 3: (((A:0.01, B:0.01):0.01, C:0.02) : $0.1, \mathrm{D}: 0.12$ ); All effective population sizes were set to 0.01 . According to coalescent theory, the level of difficulty for recovering the true species tree is determined by the ratio of internode lengths
$\boxtimes 10$ species $⿴ 囗 20$ species $\mathbb{\otimes} 30$ species


Figure 3. The branch score distance versus the number of genes. The branch score distance between the estimated species tree and the true species tree appears to converge toward 0 as the number of genes increases, but the convergence rate is negatively correlated with the number of species.
and the corresponding effective population size. The internode lengths for the population (AB) in Tree 1, Tree 2, and Tree 3 were $0.001,0.005$, and 0.1 , respectively. Hence, Tree 1 was the most difficult and Tree 3 was the easiest scenario to recover the true species tree, in terms of isometry with the contained gene trees. We used MCMCoal to generate 2, 4, 6, and 20 gene trees for each species tree. DNA sequences of 300 bp and 1000 bp were simulated for each gene tree under the Jukes-Cantor model and used as the data to estimate the species tree using BEST. The simulation and estimation was repeated 10 times for each species tree and the average posterior probability of the true species tree reported (Fig. 4). Clearly, the posterior probability of the true species tree converges to 1 as the number of genes increases (Fig. 4). However, the convergence rate depends on the true species tree. The posterior probabilities of Tree 2 and Tree 3 are already close to 1 with only six genes whereas Tree 1 required about 20 genes to reach an estimated probability of 1 . In addition, the proportion of trials yielding the true species tree was also $100 \%$ for Tree 3 when the number of genes was just 2 (Fig. 5). For Tree 2, the BEST method was able to correctly estimate the true species tree in $80 \%$ of the trials when the number of genes was six. The worst scenario was with Tree 1, in which BEST recovered the true species tree in only $50 \%$ of the trials when the number of genes was six. As


Figure 4. The posterior probability of the true species tree versus the number of genes. Three species trees are specified; Tree 1: (((A:0.01, B:0.01):0.001, C:0.011):0.1, D:0.111); Tree 2: (((A:0.01, B:0.01):0.005, C: 0.015):0.1, D:0.115); Tree 3: (((A:0.01, B:0.01):0.01, C:0.02) : 0.1, D:0.12); The program MCMCcoal (Rannala and Yang 2003) was used to generate $2,4,6$ and 20 gene trees for each species tree. DNA sequences of 300 bp and 1000 bp were simulated for each gene tree and then used as the data to estimate the species tree using BEST. The markers on each line represent the average posterior probabilities across 10 repeats and the vertical line around the marker represents the standard error. The dashed lines are the posterior probabilities for sequences of length 1000 bp. The solid lines are the posterior probabilities for sequences of length 300 bp.


Figure 5. The proportion of trials yielding the true species tree. The DNA sequences were generated from each of the three prespecified species trees; Tree 1: (((A:0.01, B:0.01):0.001, C:0.011):0.1, D:0.111); Tree 2: (((A:0.01, B:0.01):0.005, C: 0.015):0.1, D:0.115); Tree 3: (((A:0.01, B:0.01):0.01, C:0.02) : 0.1, D:0.12); The program MCMCcoal (Rannala and Yang 2003) was used to generate 2, 4, 6, and 20 gene trees for each species tree. DNA sequences of 300 bp and 1000 bp were simulated for each gene tree. The simulated sequences were analyzed by BEST to estimate the species tree. The simulation and estimation was repeated 10 times for each species tree. The dashed lines represent the proportion of trials yielding the true species tree for sequences of length 1000 bp whereas the solid lines represent the proportion of trials yielding the true species tree for sequences of length 300 bp .
the sequence length increased to 1000 bp , the average posterior probabilities of the species trees (Fig. 4) and the proportion of trials yielding the true species tree (Fig. 5) became higher than those estimated by sequences of length 300 bp , indicating that longer sequences from multiple genes are essential for accurately estimating the species tree. However, increasing sequence length alone is unable to make the Bayesian estimate converge to the true species tree. For four genes, the posterior probability of Tree 1 was 0.54 even though the sequence length had increased to 1000 bp . The discrepancy of the branch lengths between the species tree estimate and the true species tree was measured by the branch score distance. Interestingly, increasing the number of genes can only slightly improve the branch length estimation, but the branch length estimation can be dramatically improved by lengthening the sequences (Fig. 6).

There are two levels of errors in the species tree estimation process: gene tree estimation error in the first MCMC algorithm and species tree estimation error in the second MCMC algorithm. Gene tree estimation error can be reduced by lengthening sequences whereas the second estimation error can be reduced by increasing the number of genes and alleles. Obtaining an accurate Bayesian estimate of the species tree (topology and branch


Figure 6. The branch score distance of the species tree estimate and the true species tree. Three true species trees were specified to simulate two sets of DNA sequences for 2,4 , and 6 genes. The sequences in the first set were 300 bp and the sequences in the second set were 1000 bp . The sequence data were analyzed by BEST to estimate the species tree. The difference between the true species tree and the species tree estimate obtained by BEST was measured by the branch score distance. The solid lines represent the branch score distance for sequences of length 300 bp and dashed lines represent the branch score distance for sequences of length 1000 bp.
length) requires that both estimation errors be small. The simulation results (Figs. 5 and 6) suggest that gene tree estimation plays an important role in species tree estimation, especially when the branch length (or divergence time) estimation is of major interest. Reducing the gene tree estimation error is critical for accurately estimating the species divergence times, which may not be effectively achieved by increasing the number of genes.

## Data Analysis <br> yeast data

The yeast dataset (Liti et al. 2006) includes DNA sequences sampled from four loci for six species of yeast: S. cerevisiae, S. paradoxus, S. bayanus, S. cariocanus, S. mikatae, and S. kudriavzevii. The original dataset included a total of 41 alleles (strains) and six genes. To speed up computation, we removed two genes and many identical or nearly identical sequences for a given gene, reducing the dataset to 22 alleles across all species for four genes. If estimating genetic diversity of extant populations is the goal, we recommend sampling a moderate number of alleles per species ( $n=7$ or 8 ; Felsenstein 2006). Whether increased sampling within species leads to more confidence in the species tree is currently unknown. In our case we reduced the yeast dataset to ease computation. In general we do not recommend deleting sequences from datasets prior to analysis; rather we recommend collecting the appropriate data and analyzing the full dataset. If dataset reduction
is required for some reason, we recommend deleting sequences at random from species within which there is random mating, rather than selectively deleting identical sequences, which would bias estimates of genetic diversity upwards. Our method assumes that there is random mating within each tip in the species tree; the yeast dataset likely violated this assumption in so far as several species appeared structured in their genetic variation. Although our nonrandom dataset reduction could incur some ascertainment bias, we believe the bias is not strong. As in the original study, we used $S$. bayanus as the outgroup. Although hybridization among yeast strains and species is detected with increasing frequency as more species and genes are investigated (Nilson-Tillgren et al. 1980; De Barro Lopes et al. 2002; Sniegowski et al. 2002; Liti et al. 2006), transfer of genes in the yeast strains investigated is known only from subtelomeric regions, and is not known to affect the genes analyzed here.

The optimal finite sites substitution model for each locus was evaluated using approximate AIC statistics in the program MrModeltest (Nylander 2004). The GTR $+\gamma$ model (Lanave et al. 1984; Zwickl and Holder 2004) was the best fit for locus 1 (NEJ1 in original study), the HKY model for locus 2 (EST2), the GTR model for locus 3 (HDF1), and the HKY $+\gamma$ model (Hasegawa et al. 1985) for locus 4 (HDF2). Gene trees were estimated independently for each locus by MrBayes with the model specified above to investigate the variability at the gene tree level. The MCMC algorithm ran for 1,000,000 generations, with every 100th tree saved. The first 100,000 generations were discarded as the burnin. A consensus tree was constructed from the estimated posterior distribution of the gene tree for each locus.

The dataset was then analyzed in the modified MrBayes program (BEST step 1) using these locus-specific models. The prior distribution of the population size was an inverse gamma distribution with $\alpha=3$ and $\beta=0.03$. The posterior distribution of the gene trees was first estimated with the approximate joint prior of gene trees across the four genes. In this MCMC, the chain ran for 10 million generations, with every 1000th gene tree saved. For each of the 10 million cycles we sampled 1000 species trees to calculate $\tilde{f}(\mathbf{G})$. The first 1,000,000 gene trees were discarded as burnin. The estimated joint posterior distribution of gene trees was then employed to reconstruct the posterior distribution of the species tree using the second MCMC as implemented in BEST, followed by importance sampling to align the first and second posterior distributions. The species tree estimation was conducted on a $2 \times$ AMD Opteron AMD Opteron $248 / 4$ GB computer. It took 80 h for the first MCMC algorithm and 3 h for the second MCMC algorithm.

At the gene tree level, all species in the dataset were reciprocally monophyletic across loci (Fig. 7); in addition, there was strong concordance among different genes for phylogenetic relationships at the species level (Liti et al. 2006). However, there was
substantial within-strain phylogenetic heterogeneity for all four genes we examined (Fig. 7).

At the level of species, the majority rule consensus tree for the estimated posterior distribution of species trees (Fig. 8) matches the topology of the tree estimated by the concatenation method performed in the original study (Liti et al. 2006), although of course the tree in the original study had multiple tips per species whereas our tree has a single tip representing each species. The tree of these yeast species is also consistent with analyses of a 106-gene single-allele dataset encompassing these species using concatenation (Rokas et al. 2003) or the single allele Bayesian method (Edwards et al. 2007). However, the results of the BEST analysis and the concatenation analysis of the multiple-allele dataset differ in the posterior probability for the clade (S. paradoxus, $S$. cariocanus, S. cerevisiae). For the BEST method, the clade had a posterior probability of 0.57 , whereas the concatenation method estimated 1.0 for the posterior probability of the clade. Although each gene tree recovered this clade in a traditional phylogenetic analysis (Liti et al. 2006), and each species was reciprocally monophyletic for all genes, analyzed singly or together, the gene trees exhibited some heterogeneity in the internal phylogenetic structure of these three species. In such instances, concatenation may overestimate the posterior probability. Additionally, estimating confidence in species tree branches is quite different from estimating confidence in gene tree branches. Confidence in branches of the estimated species tree-the tree containing the gene trees-is likely to be a function of the number of genes as well as the confidence in each of these genes. Thus, having just a few concordant and well-supported gene trees may not be enough to guarantee confidence in the species tree, although more research in this area is needed. We are currently conducting simulations to compare the posterior probabilities of gene trees and species trees analyzed by traditional and species tree approaches to better understand the relationships between these two measures of phylogenetic confidence.

## MANAKIN (MANACUS) DATA

Manakins (Pipridae) are a family of lekking Neotropical birds, and the genus Manacus is composed of four allospecies distributed in lowland forests from southern Mexico to Brazil. We analyzed a dataset (Brumfield et al. in press), consisting of five nuclear loci from a total of 40 Manacus chromosomes, plus one allele from an outgroup, a related manakin genus Chiroxiphia pareola. Brumfield et al. (in press) treated M. manacus populations that are isolated east and west of the Andes as distinct, resulting in a total of five ingroup species-M. candei ( $n=8$ alleles), M. vitellinus ( $n=6$ ), M. aurantiacus ( $n=4$ ), M. manacus (west of Andes; $n=$ 8 ), and M. manacus (east of Andes; $n=14$ ).

The gene trees for each locus were estimated independently by MrBayes with the GTR $+\gamma$ model selected by Modeltest


Figure 7. The gene tree estimate for each locus in the yeast dataset. The gene trees were estimated by MrBayes independently for locus NEJ1 (A), EST2 (B), HDF1 (C), and HDF2 (D). Multiple alleles are present in each species. Each allele is named after the species it belongs to using three letters, as follows: $\mathrm{Sce}=$ S. cervisciae, $\mathrm{Spa}=\mathrm{S}$. paradoxus, $\mathrm{Sba}=\mathrm{S}$. bayanus, $\mathrm{Sca}=\mathrm{S}$. cariocanus, $\mathrm{Smi}=S$. mikatae, and $\mathbf{S k u}=S$. kudriavzevii. Posterior probabilities greater than or equal to 0.95 are presented.
(Posada and Crandall 1998; Posada and R. 2004). The MCMC algorithm ran for 1,000,000 generations and we saved every 100th tree. The first 100,000 generations were discarded as burnin. A consensus tree was constructed from the estimated distribution of the gene tree. The consensus tree was used as the gene tree estimate. The estimated gene trees were not well resolved and all five loci exhibited a lack of reciprocal monophyly for every ingroup species (Fig. 9). Thus, compared to the yeast dataset, the topology of the manakin species tree is not apparent upon casual inspection of the five gene trees.

To estimate the species tree, the dataset was analyzed in the modified MrBayes with the GTR $+\gamma$ model for each locus. The
prior distribution of the population size was inverse gamma with $\alpha=3$ and $\beta=0.03$. The posterior distribution of gene trees was estimated with the approximate joint prior of gene trees across the five genes. The first MCMC algorithm ran for 20 million generations and for each of these cycles we sampled 1000 species trees to calculate $\tilde{f}(\mathbf{G})$. Because we generated more gene tree vectors (20 million) than were needed, the first 10 million trees were discarded as burnin. The estimated joint posterior distribution of gene trees was then employed to reconstruct the posterior distribution of the species tree using the second MCMC algorithm as implemented in BEST, followed by importance sampling. It took 330 h for the first MCMC algorithm and 10 h for the second MCMC algorithm.


Figure 8. The species tree estimate for the yeast dataset.

The majority rule consensus tree of the estimated posterior distribution (Fig. 10) was well resolved and consisted of two main clades-(M. candei, M. aurantiacus) and (M. manacusWest, M. manacusEast, M. vitellinus). This topology agrees with a species tree for the genus estimated from isozymes (Brumfield and Braun 2001). This example is intriguing because the confidence in the species tree is quite high despite the fact that all gene trees differed from one another, and none of the ingroup species formed monophyletic groups at the level of alleles (Brumfield et al. 2007). This contrasts with the yeast example and shows that the relationship between concordance of gene trees with respect to species and confidence in the species tree may be complex.

## Discussion

Estimation of species trees can be improved by the use of molecular data with multiple alleles and multiple loci, yet in many situations (Maddison and Knowles 2006) the parameter of interest is a phylogenetic tree with branch lengths in which each tip represents a single species. DNA sequence data containing multiple alleles and multiple loci have multiple complex sources of genetic variation and error that must be addressed by analytic tools for estimating the phylogeny of the constituent species. Gene trees themselves can be challenging to estimate when branch lengths
are short and sequence data are limited. In addition, it is now well appreciated that gene tree heterogeneity is ubiquitous; because of coalescent variance, virtually any speciation scenario, whether it includes long or short branches in the species tree, will always have some heterogeneity in gene trees, if not in gene tree topology then in gene tree branch lengths. Finally, whereas branches of gene trees are likely to vary within species, again due to the coalescent, if the branches of the species tree are long, these same gene trees will show substantial consistency in deeper branches of the species tree.

Each of these sources of variance has associated errors in estimation, but most current tree reconstruction methods lack the capability to incorporate all these sources of variance. The concatenation method, the most popular approach for inferring phylogenetic relationships, is unable to accommodate variation among loci because it assumes that all gene trees have the same topology. A model that can explain these variations must involve both gene trees and species trees as two correlated quantities. It could be extremely difficult for nonmodel-based methods to construct a reasonable function to estimate the species tree for multiple alleles and multiple loci data. Furthermore, nonmodel-based methods are often difficult to generalize themselves when more complexities appear in the data.

As analytic tools, model-based approaches have substantial promise for analyzing high-dimensional sequence data. The Bayesian hierarchical model we propose, although still at an early stage, shows promise in its ability to handle datasets with strong interlocus concordance (the yeast dataset) as well as datasets with substantial heterogeneity among genes and incomplete coalescence among species (manakin dataset). However, as with any statistical model, it is important to develop an understanding of how deviations from model assumptions affect the conclusions drawn. Our Bayesian hierarchical model assumes that incomplete lineage sorting is the sole source of incongruence among gene trees and between gene trees and the species tree. However, conflicts among gene trees and between gene and species trees may be caused by other factors such as horizontal gene transfer, hybridization, and gene duplication/gene loss (Maddison 1997). A more realistic model demands the incorporation of these factors when estimating the species tree (Linz et al. 2007).

Previous studies suggest that introgression could have influenced the Manacus dataset (Brumfield et al. 2001, in press), which would violate BEST's assumption that no gene flow occurred after speciation. Despite this possibility, BEST still delivered reasonable species tree estimates congruent with other data. This suggests the BEST method may be robust to potential assumption violations. Some of the earliest phylogenetic studies analyzed multilocus datasets from human populations, despite known introgression among these populations (Cavalli-Sforza and Edwards 1964). Even today, phylogenetic trees of populations or species


Figure 9. The gene tree estimates for the Manacus dataset. The posterior distribution of the gene trees was estimated using MrBayes independently for each locus; $\beta$-actin intron 3 (A), $\beta$-fibrinogen intron 7 (B), ornithine decarboxylase introns 6 and 7 (C), rhodopsin intron 2 (D), and transforming growth factor $\beta 2$ intron $2(E)$. The consensus tree was constructed from the estimated posterior distribution of the gene tree. Posterior probabilities greater than or equal to 0.95 were indicated by an asterisk next to the specific branch. Outgroups were removed from the consensus trees. The first three or four letters of the species were used as the abbreviation of the species. The following number represented the allele sampled from each species.
are sometimes constructed from population statistics such as $F_{\mathrm{ST}}$, even when hybridization is suspected. Hybridization may not be a problem for estimating topologies of species trees when it occurs between sister species; in such situations, the sister species will still be resolved as such, albeit with shorter branch lengths than if no hybridization occurred. If the introgressing gene regions only reflect a small percentage of the entire genome (Rieseberg et al. 1999), then it may be unlikely that the gene trees were influenced by hybridization. What is certain is that reticulate patterns of evo-
lution are quite prevalent and easily detectable once multiple loci are examined (Bensch et al. 2006). Clearly, the effects of hybridization, horizontal transfer, migration, gene duplication/loss on the BEST model need to be further studied. Equally important will be the development of methods for determining whether hybridization or incomplete lineage sorting is the cause of gene tree discordances as a means of choosing the appropriate model for analysis. Several approaches, including method-of-moments, maximum likelihood and approximate Bayesian approaches, seem


Figure 10. The estimate of the species tree for the Manacus dataset.
promising in this regard (Wakeley 1996; Beaumont and Rannala 2004; Hey and Nielsen 2007).

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## LITERATURE CITED

Alfaro, M. E., and M. T. Holder. 2006. The posterior and the prior in Bayesian phylogenetics. Annu. Rev. Ecol. Evol. Syst. 37:19-42.
Beaumont, M. A., and B. Rannala. 2004. The Bayesian revolution in genetics. Nat. Rev. Genet. 5:251-261.
Beerli, P. 2006. Comparison of Bayesian and maximum likelihood inference of population genetic parameters. Bioinformatics 22:341-345.
Bensch, S., D. E. Irwin, J. H. Irwin, L. Kvist, and S. Akesson. 2006. Conflicting patterns of mitochondrial and nuclear DNA diversity in Phylloscopus warblers. Mol. Ecol. 15:161-171.
Brumfield, R. T., and M. J. Braun. 2001. Phylogenetic relationships in bearded manakins (Pipridae; Manacus) indicate that male plumage color is a misleading taxonomic marker. Condor 103:248-258.
Brumfield, R. T., R. W. Jernigan, D. B. McDonald, and M. J. Braun. 2001. Evolutionary implications of divergent clines in an avian (Manacus; Aves) hybrid zone. Evolution 55:2070-2087.

Brumfield, R. T., L. Liu, D. E. Lum, and S. V. Edwards. in press. Comparison of species tree methods for reconstructing the phylogeny of bearded manakins (Aves: Pipridae, Manacus) from multilocus sequence data. Syst. Biol.
Bull, J. J., J. P. Huelsenbeck, C. W. Cunningham, D. L. Swofford, and P. J. Waddell. 1993. Partitioning and combining data in phylogenetic analysis. Syst. Biol. 42:384-397.
Cavalli-Sforza, L. L., and A. W. F. Edwards. 1964. Analysis of human evolution. Proc. XI Int. Congress Genet. 3:923-933.
Cranston, K. A., and B. Rannala. 2007. Summarizing a posterior distribution of trees using agreement subtrees. Syst. Biol. 56:578-590.
De Barro Lopes, M., J. R. Bellon, N. J. Shirley, and P. F. Ganter. 2002. Evidence for multiple interspecific hybridization in Saccharomyces sensu stricto species. FEM Yeast Res. 1:323-331.
de Queiroz, A. 1993. For consensus (sometimes). Syst. Biol. 42:368-372.
de Queiroz, A., M. J. Donoghue, and J. Kim. 1995. Separate versus combined analysis of phylogenetic evidence. Annu. Rev. Ecol. Syst. 26:657-681.
Degnan, J. H., and N. A. Rosenberg. 2006. Discordance of species trees with their most likely gene trees. PLoS Genet. 2:762-768.
Degnan, J. H., and L. A. Salter. 2005. Gene tree distributions under the coalescent process. Evolution 59:24-37.
Edwards, S. V., L. Liu, and D. K. Pearl. 2007. High-resolution species trees without concatenation. Proc. Natl. Acad. Sci. USA 104:5936-5941.
Felsenstein, J. 2004. Inferring phylogenies. Sinauer Associates, Sunderland, MA.
-. 2006. Accuracy of coalescent likelihood estimates: do we need more sites, more sequences, or more loci? Mol. Biol. Evol. 23:691-700.
Funk, D. J., and K. E. Omland. 2003. Species-level paraphyly and polyphyly: frequency, causes, and consequences, with insights from animal mitochondrial DNA. Annu. Rev. Ecol. Evol. Syst. 34:397-423.
Hasegawa, M., H. Kishino, and T. A. Yano. 1985. Dating of the human ape splitting by a molecular clock of Mitochondrial-DNA. J. Mol. Evol. 22:160-174.
Hastings, W. K. 1970. Monte-Carlo sampling methods using Markov Chains and their applications. Biometrika 57:97-109.
Hey, J., and R. Nielsen. 2004. Multilocus methods for estimating population sizes, migration rates and divergence time, with applications to the divergence of Drosophila pseudoobscura and $D$. persimilis. Genetics 167:747-760.

- 2007. Integration within the Felsenstein equation for improved Markov Chain Monte Carlo methods in population genetics. Proc. Natl. Acad. Sci. USA 104:2785-2790.
Huelsenbeck, J. P., and F. Ronquist. 2001. MRBAYES: Bayesian inference of phylogenetic trees. Bioinformatics 17:754-755.
Huelsenbeck, J. P., D. L. Swofford, C. W. Cunningham, J. J. Bull, and P. J. Waddell. 1994. Is character weighting a panacea for the problem of data heterogeneity in phylogenetic analysis. Syst. Biol. 43:288-291.
Kingman, J. F. C. 1982. On the genealogy of large populations. Stoch. Proc. Appl. 13:235-248.
- 2000. Origins of the coalescent: 1974-1982. Genetics 156:14611463.

Kuhner, M. K., and J. Felsenstein. 1994. A simulation comparison of phylogeny algorithms under equal and unequal evolutionary rates. Molecular Biology and Evolution 11:459-468.
Lanave, C., G. Preparata, C. Saccone, and G. Serio. 1984. A new method for calculating evolutionary substitution rates. J. Mol. Evol. 20:86-93.
Linz, S., A. Radtke, and A. von Haeseler. 2007. A likelihood framework to measure horizontal gene transfer. Mol. Biol. Evol. 24:1312-1319.
Liti, G., D. B. Barton, and E. J. Louis. 2006. Sequence diversity, reproductive isolation and species concepts in Saccharomyces. Genetics 174:839850.

Liu, L. 2006. Reconstructing posterior distributions of a species phylogeny using estimated gene tree distributions. Pp. 46-49. in Ph.D dissertation. Department of Statistics. The Ohio State Univ., Columbus.
Liu, L., and D. K. Pearl. 2007. Species trees from gene trees: reconstructing Bayesian posterior distributions of a species phylogeny using estimated gene tree distributions. Syst. Biol. 56:504-514.
Maddison, W. P. 1997. Gene trees in species trees. Syst. Biol. 46:523-536.
Maddison, W. P., and L. L. Knowles. 2006. Inferring phylogeny despite incomplete lineage sorting. Syst. Biol. 55:21-30.
Maddison, W. P., and D. R. Maddison. 2006. Mesquite: a modular system for evolutionary analysis. Version 1.12.
Mossel, E., and S. Roch. 2007. Incomplete lineage sorting: consistent phylogeny estimation from multiple Loci. Available at http://arxiv.org/ abs/0710.0262 [Accessed November 2, 2007].
Nee, S., R. M. May, and P. H. Harvey. 1994. The reconstructed evolutionary process. Philos. Trans. R. Soc. Lond. B 344:305-311.
Newton, M. A., A. E. Raftery, A. C. Davison, M. Bacha, G. Celeux, B. P. Carlin, P. Clifford, C. Lu, M. Sherman, M. A. Tanner, et al. 1994. Approximate Bayesian-inference with the weighted likelihood bootstrap. J. R. Stat. Soc. B 56:3-48.
Nielsen, R., and M. Slatkin. 2000. Likelihood analysis of ongoing gene flow and historical association. Evolution 54:44-50.
Nilson-Tillgren, T., J. G. L. Petersen, S. Holmberg, and M. C. Kielland-Brandt. 1980. Transfer of chromosome III during Kar mediated cytoduction in yeast Carlsberg. Res. Commun. 45:113-117.
Nylander, J. A. A. 2004. MrModeltest v2. Evolutionary Biology Centre, Uppsala Univ.
Nylander, J. A. A., F. Ronquist, J. P. Huelsenbeck, and J. L. Nieves-Aldrey. 2004. Bayesian phylogenetic analysis of combined data. Syst. Biol. 53:47-67.
Page, R. D. M. 1998. GeneTree: comparing gene and species phylogenies using reconciled trees. Bioinformatics 14:819-820.
Page, R. D. M., and M. A. Charleston. 1997. From gene to organismal phylogeny: reconciled trees and the gene tree species tree problem. Mol. Phylogenet. Evol. 7:231-240.
Posada, D., and K. A. Crandall. 1998. MODELTEST: testing the model of DNA substitution. Bioinformatics 14:817-818.

Posada, D., and T. R. Buckley. 2004. Model selection and model averaging in phylogenetics: advantages of the AIC and Bayesian approaches over likelihood ratio tests. Syst. Biol. 53:793-808.
Rannala, B., and Z. H. Yang. 2003. Bayes estimation of species divergence times and ancestral population sizes using DNA sequences from multiple loci. Genetics 164:1645-1656.
Rieseberg, L. H., J. Whitton, and K. Gardner. 1999. Hybrid zones and the genetic architecture of a barrier to gene flow between two sunflower species. Genetics 152:713-727.
Rokas, A., B. L. Williams, N. King, and S. B. Carroll. 2003. Genome-scale approaches to resolving incongruence in molecular phylogenies. Nature 425:798-804.
Ronquist, F., and J. P. Huelsenbeck. 2003. MrBayes 3: Bayesian phylogenetic inference under mixed models. Bioinformatics 19:15721574.

Slowinski, J. B., A. Knight, and A. P. Rooney. 1997. Inferring species trees from gene trees: a phylogenetic analysis of the elapidae (Serpentes) based on the amino acid sequences of venom proteins. Mol. Phylogenet. Evol. 8:349-362.
Sniegowski, P. D., P. G. Dombrowski, and E. Fingerman. 2002. Saccharomyces cerevisiae and Saccharomyces paradoxus coexist in a natural woodland site in North America and display different levels of reproductive isolation from European conspecifics. FEMS Yeast Res. 1:299306.

Wakeley, J. 1996. The variance of pairwise nucleotide differences in two populations with migration. Theor. Popul. Biol. 49:39-57.
Whelan, S., and N. Goldman. 2001. A general empirical model of protein evolution derived from multiple protein families using a maximumlikelihood approach. Mol. Biol. Evol. 18:691-699.
Wiens, J. J. 1998. Combining data sets with different phylogenetic histories. Syst. Biol. 47:568-581.
Zwickl, D. J., and M. T. Holder. 2004. Model parameterization, prior distributions, and the general time-reversible model in Bayesian phylogenetics. Syst. Biol. 53:877-888.

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